ANNEX II

APPROVAL OF ESTABLISHMENTS

[F1CHAPTER III

DISEASE SURVEILLANCE PROGRAMME

Without prejudice to health measures and to Articles 16 and 17, disease surveillance programmes must, as a minimum, comprise surveillance of the infections and species listed in Sections A to D.

A. *Salmonella* Pullorum[(1)], *Salmonella* Gallinarum[(2)] and *Salmonella arizonae*[(3)] infections

1. Species concerned

(a) *Salmonella* Pullorum and *Salmonella* Gallinarum: fowls, turkeys, guinea fowls, quails, pheasants, partridges and ducks.

(b) *Salmonella arizonae*: turkeys.

2. Disease surveillance programme

(a) Serological and/or bacteriological tests must be used to determine whether an infection is present[(4)].

(b) Samples for testing must be taken, as appropriate, from blood, embryos that fail to hatch (namely embryos dead-in-shell), second grade chicks, meconium, post mortem tissues, especially liver, spleen, ovary/oviduct and ileo-caecal junction[(5)].

(c) Direct enrichment in Selenite-cysteine broth for faecal/meconium and intestinal samples is to be used. Non-selective pre-enrichment followed by selective enrichment in soya based Rappaport-Vassiliadis (RVS) broth or Müller-Kauffmann Tetrathionate-novobiocin broth (MKTTn) may be used for samples (such as embryos dead-in-shell) where competing flora is expected to be minimal[(6)],[(7)].

(d) When blood samples are taken from a flock for serological testing for *Salmonella* Pullorum and *Salmonella* Gallinarum or *Salmonella arizonae*, the prevalence of infection in the Member State concerned and its past incidence in the establishment must be allowed for in determining the number of samples to be taken. However, a statistically valid number of samples for serological and/or bacteriological testing to detect infection must always be taken.

(e) Flocks must be inspected during each laying period at the best time for detecting the disease in question.

(f) Samples for bacteriological testing must not be taken from poultry or eggs that have been treated with antimicrobial medicinal products during the 2 to 3 weeks prior to testing.

(g) Detection techniques must be capable of differentiating serological responses to *Salmonella* Pullorum and *Salmonella* Gallinarum infection from serological responses due to the use of *Salmonella* Enteritidis vaccine, where this vaccine is used[(8)]. Such vaccination must therefore not be used if serological monitoring is to be used. If

vaccination has been used, bacteriological testing must be used, but the confirmation method used must be capable of differentiating live vaccinal strains from field strains.

B. *Mycoplasma gallisepticum* and *Mycoplasma meleagridis* infections

1. Species concerned

(a) *Mycoplasma gallisepticum*: fowls and turkeys.

(b) *Mycoplasma meleagridis*: turkeys.

2. Disease surveillance programme

(a) The presence of infection must be tested by validated serological and/or bacteriological and/or molecular tests. The presence of air sacculitis lesions in day-old chicks and turkey poults suggests that a *Mycoplasma* infection is present and must be investigated.

(b) Samples for testing for the presence of *Mycoplasma* infection must be taken, as appropriate, from blood, day-old chicks and turkey poults, sperm, or swabs taken from the trachea, the choanae, cloaca or air sacs and in particular for the detection of *Mycoplasma meleagridis* samples must be taken from oviduct and penis of turkeys.

(c) Tests for detecting *Mycoplasma gallisepticum* or *Mycoplasma meleagridis* must be performed on a representative sample in order to allow continuous surveillance of the infection during rearing and laying, namely just before the start of laying and every 3 months thereafter.

C. Results and measures to be taken

If there are no reactors, the test must be deemed to be negative. Otherwise, the flock must be deemed to be suspect and the measures set out in Chapter IV must be applied to it.

D. In the case of holdings which consist of two or more separate production units, the competent veterinary authority may derogate from the measures set out in point 3(b) of Chapter IV required for restoring of approval as regards healthy production units on a holding where the infection is present provided that the authorised veterinarian has confirmed that the structure and size of those production units and the operations carried out there are such that the production units provide completely separate facilities for housing, keeping and feeding, so that the disease in question cannot spread from one production unit to another.**]**

Textual Amendments

F1 Substituted by Commission Decision of 1 April 2011 amending Annexes II to IV to Council Directive 2009/158/EC on animal health conditions governing intra-Community trade in, and imports from third countries of, poultry and hatching eggs (notified under document C(2011) 2068) (Text with EEA relevance) (2011/214/EU).

(1) **[***F1* *Salmonella* Pullorum means *Salmonella enterica* subspecies *enterica* serovar Gallinarum biochemical variant (biovar) Pullorum.**]**

(2) **[***F1* *Salmonella* Gallinarum means *Salmonella enterica* subspecies *enterica* serovar Gallinarum biochemical variant (biovar) Gallinarum.**]**

(3) **[***F1* *Salmonella arizonae* means *Salmonella enterica* subspecies *arizonae* serogroup K (O18) arizonae.**]**

(4) **[**F1Note that serological testing in avian species other than fowls may sometimes result in an unacceptable proportion of false-positive reactions.**]**

(5) **[**F1**[**Note that environmental samples are generally not suitable for reliable detection of *Salmonella pullorum* and *Salmonella gallinarum*, but are suitable for *Salmonella arizonae.**]]***

(6) **[**F1Note that direct plating of aseptically collected tissues on to a minimally selective agar, such as MacConkey agar, is also useful for diagnosis.**]**

(7) **[***F1* ***[****Salmonella pullorum* and *Salmonella gallinarum* do not readily grow in the modified semi-solid Rappaport Vassiliadis (MSRV) medium that is used for monitoring of zoonotic *Salmonella* spp. in the Union, but it is suitable for *Salmonella arizonae.**]]***

(8) **[**F1Note that there is currently no test that can differentiate between the response to *Salmonella* Pullorum and *Salmonella* Gallinarum infection and vaccination for this serotype.**]**

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