Commission Implementing Regulation (EU) 2016/1240 of 18 May 2016 laying down rules for the application of Regulation (EU) No 1308/2013 of the European Parliament and of the Council with regard to public intervention and aid for private storage (Text with EEA relevance)

[F1Appendix

Textual Amendments

F1 Inserted by Commission Implementing Regulation (EU) 2018/150 of 30 January 2018 amending Implementing Regulation (EU) 2016/1240 as regards methods for the analysis and quality evaluation of milk and milk products eligible for public intervention and aid for private storage.

METHOD FOR THE DETECTION OF COW'S MILK AND CASEINATE IN CHEESES FROM EWE'S MILK, GOAT'S MILK OR BUFFALO MILK OR MIXTURES OF EWE'S MILK, GOAT'S MILK AND BUFFALO MILK

1. SCOPE

Detection of cow's milk and caseinate in cheeses made from ewe's milk, goat's milk, buffalo milk or mixtures of ewe's, goat's and buffalo milk by isoelectric focusing of γ-caseins after plasminolysis.

2. FIELD OF APPLICATION

The method is suitable for sensitive and specific detection of native and heat-treated cow's milk and caseinate in fresh and ripened cheeses made from ewe's milk, goat's milk, buffalo milk or mixtures of ewe's, goat's and buffalo milk. It is not suitable for the detection of milk and cheese adulteration by heat-treated bovine whey protein concentrates.

3. PRINCIPLE OF THE METHOD

3.1. **Isolation of caseins from cheese and the reference standards**

3.2. **Dissolving of the isolated caseins and submitting to plasmin (EC.3.4.21.7) cleavage**

3.3. **Isoelectric focusing of plasmin-treated caseins in the presence of urea and staining of proteins**

3.4. **Evaluation of stained γ_3 and γ_2 -casein patterns (evidence of cow's milk) by comparison of the pattern obtained from the sample with those obtained in the same gel from the reference standards containing 0 % and 1 % cow's milk.**

4. REAGENTS

Unless otherwise indicated, analytical grade chemicals shall be used. Water shall be double-distilled or of equivalent purity.

Note: The following details apply to laboratory prepared polyacrylamide gels containing urea, of dimensions 265 × 125 × 0,25 mm. Where other sizes and types of gel are used, the separation conditions may have to be adjusted.

Isoelectric focusing

4.1. **Reagents for production of the urea containing polyacrylamide gels**

4.1.1. *Stock gel solution*

Dissolve:

4,85 g acrylamide
0,15 g N, N′-methylene-bis-acrylamide (BIS)
48,05 g urea

15,00 g glycerol (87 % w/w),

in water and make up to 100 ml and store in a brown glass bottle in the refrigerator.

Note: A commercially available pre-blended acrylamide/BIS solution may be used in preference to the quoted fixed weights of the neurotoxic acrylamides. Where such a solution contains 30 % w/v acrylamide and 0,8 % w/v BIS, a volume of 16,2 ml shall be used for the formulation instead of the fixed weights. The shelf life of the stock solution is a maximum of 10 days; if its conductivity is more than 5 μS, de-ionize by stirring with 2 g Amberlite MB-3 for 30 minutes, then filter through a 0,45 μm membrane.

4.1.2. *Gel solution*

Prepare a gel solution by mixing additives and ampholytes (*) with the stock gel solution (see 4.1.1).

9,0 ml stock solution
24 mg β-alanine
500 μl ampholyte pH 3,5-9,5
250 μl ampholyte pH 5-7
250 μl ampholyte pH 6-8

Mix the gel solution and de-gas for two to three minutes in an ultrasonic bath or in vacuum.

Note: Prepare the gel solution immediately prior to pouring it (see 6.2).

4.1.3. *Catalyst solutions*

4.1.3.1. N, N, N′ N′ — tetramethylethylenediamine (Temed)

4.1.3.2. 40 % w/v ammonium persulphate (PER):

Dissolve 800 mg PER in water and make up to 2 ml.

Note: Always use freshly prepared PER solution.

4.2. **Contact fluid**

Kerosene or liquid paraffin

4.3. **Anode solution**

Dissolve 5,77 g phosphoric acid (85 % w/w) in water and dilute to 100 ml.

4.4. **Cathode solution**

Dissolve 2,00 g sodium hydroxide in water and dilute to 100 ml with water.
Sample preparation

4.5. **Reagents for protein isolation**

4.5.1. *Dilute acetic acid (25,0 ml of glacial acetic acid made up to 100 ml with water)*

4.5.2. *Dichloromethane*

4.5.3. *Acetone*

4.6. **Protein dissolving buffer**

Dissolve

5,75 g glycerol (87 % w/w)
24,03 g urea
250 mg dithiothreitol,

in water and make up to 50 ml

Note: Store in a refrigerator, maximum shelf-life one week.

4.7. **Reagents for plasmin cleavage of caseins**

4.7.1. *Ammonium carbonate buffer*

Titrate a 0,2 mol/l ammonium hydrogencarbonate solution (1,58 g/100 ml water) containing 0,05 mol/l ethylenediaminetetraacetic acid (EDTA, 1,46 g/100 ml with a 0,2 mol/l ammonium carbonate solution (1,92 g/100 ml water) containing 0,05 mol/l EDTA to pH 8.

4.7.2. *Bovine plasmin (EC. 3.4.21.7), activity at least 5 U/ml*

4.7.3. *ε-Aminocaproic acid solution for enzyme inhibition*

Dissolve 2,624 g ε-aminocaproic acid (6 amino-n-hexanoic acid) in 100 ml of 40 % (v/v) ethanol.

4.8. **Standards**

4.8.1. *Certified reference standards of a mixture of renneted ewe's and goat's skimmed milk containing 0 % and 1 % of cow's milk are available from the Commission's Institute for Reference Materials and Measurements, B-2440 Geel, Belgium*

4.8.2. *Preparation of laboratory interim-standards of buffalo's renneted milk containing 0 % and 1 % of cow's milk*

Skimmed milk is prepared by centrifuging of either buffalo or bovine raw bulk milk at 37 °C at 2 500 g for 20 minutes. After cooling the tube and contents rapidly to 6 to 8 °C, the upper fat layer is removed completely. For the preparation of the 1 % standard add 5,00 ml of bovine skimmed milk to a 495 ml of buffalo's skimmed milk in a 1 l beaker, adjust the pH to 6,4 by the addition of dilute lactic acid (10 % w/v). Adjust the temperature to 35 °C and add 100 µl of calf rennet (rennet activity 1: 10 000, c. 3 000 U/ml), stir for 1 minute and then leave the beaker covered with an aluminium foil at 35 °C for one hour to allow formation of the curd. After the curd has formed, the whole renneted milk is freeze-dried without prior homogenization or draining of the whey. After freeze-drying it is finely ground to produce a homogeneous powder. For the preparation of the 0 % standard, carry out the same procedure using genuine buffalo skimmed milk. The standards shall be stored at – 20 °C.

Note: It is advisable to check the purity of the buffalo milk by isoelectric focusing of the plasmin-treated caseins before preparation of the standards.

Reagents for protein staining

4.9. **Fixative**

Dissolve 150 g trichloroacetic acid in water and make up to 1 000 ml.

4.10. **Destaining solution**

Dilute 500 ml methanol and 200 ml glacial acetic acid to 2 000 ml with distilled water.

Note: Prepare the destaining solution fresh every day; it can be prepared by mixing equal volumes of stock solutions of 50 % (v/v) methanol and 20 % (v/v) glacial acetic acid.

4.11. **Staining solutions**

4.11.1. *Staining solution (stock solution 1)*

Dissolve 3,0 g Coomassie Brilliant Blue G-250 (C.I. 42655) in 1 000 ml 90 % (v/v) methanol using a magnetic stirrer (approximately 45 minutes), filter through two medium-speed folded filters.

4.11.2. *Staining solution (stock solution 2)*

Dissolve 5,0 g copper sulphate pentahydrate in 1 000 ml 20 % (v/v) acetic acid.

4.11.3. *Staining solution (working solution)*

Mix together 125 ml of each of the stock solutions (4.11.1, 4.11.2) immediately prior to staining.

Note: The staining solution should be prepared on the day that it is used.

5. EQUIPMENT

5.1. **Glass plates (265 × 125 × 4 mm); rubber roller (width 15 cm); levelling table**

5.2. **Gel carrier sheet (265 × 125 mm)**

5.3. **Covering sheet (280 × 125 mm). Stick on strip of adhesive tape (280 × 6 × 0,25 mm) to each long edge (see Figure 1)**

5.4. **Electrofocusing chamber with cooling plate (e.g. 265 × 125 mm) and suitable power supply (≥ 2,5 kV) or automatic electrophoresis device**

5.5. **Circulation cryostat, thermostatically controlled at 12 ± 0,5 °C**

5.6. **Centrifuge, adjustable to 3 000 g**

5.7. **Electrode strips (≥ 265 mm long)**

5.8. **Plastic dropping bottles for the anode and cathode solutions**

5.9. **Sample applicators (10 × 5 mm, viscose or low protein-adsorption filter paper)**

5.10. **Stainless steel or glass staining and destaining dishes (e.g. 280 × 150 mm instrument trays)**

5.12. **Adjustable rod homogenizer (10 mm shaft diameter), rpm range 8 000 to 20 000**

5.13. **Magnetic stirrer**

5.14. **Ultrasonic bath**

5.15. **Film welder**

5.16. **25 µl micropipettes**

5.17. **Vacuum concentrator or freeze-dryer**

5.18. **Thermostatically controlled water bath adjustable to 35 and 40 ± 1 °C with shaker**

5.19. **Densitometer equipment reading at λ = 634 nm**

6. PROCEDURE

6.1. Sample preparation

6.1.1. *Isolation of caseins*

Weigh the amount equivalent to 5 g dry mass of cheese or the reference standards into a 100 ml centrifuge tube, add 60 ml distilled water and homogenize with a rod homogenizer (8 000 to 10 000 rpm). Adjust to pH 4,6 with dil. acetic acid (4.5.1) and centrifuge (5 minutes, 3 000 g). Decant the fat and whey, homogenize the residue at 20 000 rpm in 40 ml distilled water adjusted to pH 4,5 with dil. acetic acid (4.5.1), add 20 ml dichloromethane (4.5.2), homogenize again and centrifuge (5 minutes, 3 000 g). Remove the casein layer that lies between the aqueous and organic phases (see Figure 2) with a spatula and decant off both phases. Rehomogenise the casein in 40 ml distilled water (see above) and 20 ml dichloromethane (4.5.2) and centrifuge. Repeat this procedure until both extraction phases are colourless (two to three times). Homogenize the protein residue with 50 ml acetone (4.5.3) and filter through a medium-speed folded filter paper. Wash the residue on the filter with two separate 25 ml portions of acetone each time and allow to dry in the air or a stream of nitrogen, then pulverize finely in a mortar.

Note: Dry casein isolates should be kept at –20 °C.

6.1.2. *Plasmin cleavage of β-caseins to intensify γ-caseins*

Disperse 25 mg of isolated caseins (6.1.1) in 0,5 ml ammonium carbonate buffer (4.7.1) and homogenize for 20 minutes by e.g. using ultrasonic treatment. Heat to 40 °C and add 10 µl plasmin (4.7.2), mix and incubate for one hour at 40 °C with continuous shaking. To inhibit the enzyme add 20 µl ε-aminoproic acid solution (4.7.3), then add 200 mg of solid urea and 2 mg of dithiothreitol.

Note: To obtain more symmetry in the focused casein bands it is advisable to freeze-dry the solution after adding the ε-aminocaproic acid and then dissolving the residues in 0,5 ml protein dissolving buffer (4.6).

6.2. Preparation of the urea containing polyacrylamide gels

With the aid of a few drops of water roll the gel carrier sheet (5.2) onto a glass plate (5.1), removing any extraneous water with paper towel or tissue. Roll the cover sheet (5.3) with spacers (0,25 mm) onto another glass plate in the same way. Lay the plate horizontally on a levelling table.

Add 10 µl Temed (4.1.3.1) to the prepared and de-aerated gel solution (4.1.2), stir and add 10 µl PER-solution (4.1.3.2), mix thoroughly and immediately pour out evenly onto the centre of the cover sheet. Place one edge of the gel carrier plate (sheet side down) on the cover sheet plate and lower it slowly so that a gel film forms between the sheets and spreads out regularly and free of bubbles (Figure 3). Carefully lower the gel carrier plate completely using a thin spatula and place three more glass plates on top of it to act as weights. After polymerization is complete (about 60 minutes) remove the gel polymerized onto the gel carrier sheet along with the cover sheet by tipping the glass plates. Clean the reverse of the carrier sheet carefully to remove gel residues and urea. Weld the gel sandwich into a film tube and store in a refrigerator (maximum six weeks).

Note: The cover sheet with the spacers can be re-used. The polyacrylamide gel can be cut to smaller sizes, recommended when there are few samples or if an automatic electrophoresis device is used (two gels, size 4,5 × 5 cm).

6.3. Isoelectric focusing

Set the cooling thermostat to 12 °C. Wipe off the reverse of the gel carrier sheet with kerosene, then drip a few drops of kerosene (4.2) onto the centre of the cooling block. Then roll the gel sandwich, carrier side down, onto it, taking care to avoid bubbles. Wipe off any excess kerosene and remove the cover sheet. Soak the electrode strips with the electrode solutions (4.3, 4.4), cut to gel length and place in the positions provided (distance of electrodes 9,5 cm).

Conditions for isoelectric focusing:

6.3.1. *Gel size 265 × 125 × 0,25 mm*

Step		Time(min.)	Voltage(V)	Current(mA)	Power(W)	Volt-hours(Vh)
1.	Pre-focusing	30	maximum 2 500	maximum 15	constant 4	c. 300
2.	Sample focusing[a]	60	maximum 2 500	maximum 15	constant 4	c. 1 000
3.	Final focusing	60	maximum 2 500	maximum 5	maximum 20	c. 3 000
		40	maximum 2 500	maximum 6	maximum 20	c. 3 000
		30	maximum 2 500	maximum 7	maximum 25	c. 3 000

a Sample application: After pre-focusing (step 1), pipette 18 µl of the sample and standard solutions onto the sample applicators (10 × 5 mm), place them on the gel at 1 mm intervals from each other and 5 mm longitudinally from the anode and press lightly. Carry out focusing using the above conditions, carefully removing the sample applicators after the 60 minutes of sample focusing.

Note: If thickness or width of the gels are changed, the values for current and power have to be suitably adjusted (e.g. double the values for electric current and power if a 265 × 125 × 0,5 mm gel is used).

6.3.2. *Example of a voltage programme for an automatic electrophoresis device (2 gels of 5,0 × 4,5 cm), electrodes without strips applied directly to the gel*

Step		Voltage	Current	Power	Temp.	Volt-hours
1.	Pre-focusing	1 000 V	10,0 mA	3,5 W	8 °C	85 Vh
2.	Sample focusing	250 V	5,0 mA	2,5 W	8 °C	30 Vh
3.	Focusing	1 200 V	10,0 mA	3,5 W	8 °C	80 Vh
4.	Focusing	1 500 V	5,0 mA	7,0 W	8 °C	570 Vh

Place sample applicator in step 2 at 0 Vh.

Remove sample applicator in step 2 at 30 Vh.

6.4. **Protein staining**

6.4.1. *Protein fixation*

Remove the electrode strips immediately after turning off the power and put the gel immediately into a staining/destaining dish filled with 200 ml fixative (4.9); leave for 15 minutes, shaking continuously.

6.4.2. *Washing and staining the gel plate*

Thoroughly drain off the fixative and wash the gel plate twice for 30 seconds each time with 100 ml destaining solution (4.10). Pour off the destaining solution and fill the dish with 250 ml staining solution (4.11.3); allow to stain for 45 minutes with gentle shaking.

6.4.3. *Destaining the gel plate*

Pour off the staining solution, wash the gel plate twice using a 100 ml destaining solution (4.10) each time, then shake with 200 ml destaining solution for 15 minutes and repeat the destaining step at least two or three times until the background is clear and uncoloured. Then rinse the gel plate with distilled water (2 × 2 minutes) and dry in the air (2 to 3 hours) or with a hairdryer (10 to 15 minutes).

Note 1: Carry out fixing, washing, staining and destaining at 20 °C. Do not use elevated temperatures.

Note 2: If more sensitive silver staining (e.g. Silver Staining Kit, Protein, Pharmacia Biotech, Code No 17-1150-01) is preferred, plasmin-treated casein samples have to be diluted to 5 mg/ml.

7. EVALUATION

Evaluation is performed by comparing the protein patterns of the unknown sample with reference standards on the same gel. Detection of cow's milk in cheeses from ewe's milk, goat's milk and buffalo milk and mixtures of ewe's, goat's and buffalo milk is done via the γ_3- and γ_2-caseins, whose isoelectric points range between pH 6,5 and pH 7,5 (Figures 4 a, b, Figure 5). The detection limit is less than 0,5 %.

7.1. **Visual estimation**

For visual evaluation of the amount of bovine milk it is advisable to adjust the concentrations of samples and standards to obtain the same level of intensity of the ovine, caprine and/or buffalo γ_2- and γ_3-caseins (see 'γ_2 E,G,B' and 'γ_3 E,G,B' in Figures 4 a, b and Figure 5). After which the amount of bovine milk (less than, equal to or greater than 1 %) in the unknown sample can be judged directly by comparing the intensity of the bovine γ_3- and γ_2-caseins (see 'γ_3 C' and 'γ_2 C' in Figures 4 a, b and Figure 5) to those of the 0 % and 1 % reference standards (ewe, goat) or, laboratory interim-standards (buffalo).

7.2. **Densitometric estimation**

If available, apply densitometry (5.19) for the determination of the peak area ratio of bovine to ovine, caprine and/or buffalo γ_2- and γ_3-caseins (see Figure 5). Compare this value to γ_2- and γ_3-casein peak area ratio of the 1 % reference standard (ewe, goat) or laboratory interim-standard (buffalo) analysed on the same gel.

Note: The method is operating satisfactorily, if there is a clear positive signal for both bovine γ_2- and γ_3-caseins in the 1 % reference standard but not in the 0 % reference standard. If not, optimize the procedure following the details of the method precisely.

A sample is judged as being positive, if both bovine γ_2- and γ_3-caseins or the corresponding peak area ratios are equal to or greater than the level of the 1 % reference standard.

8. REFERENCES

Addeo F., Moio L., Chianese L., Stingo C., Resmini P., Berner I, Krause I., Di Luccia A., Bocca A.: Use of plasmin to increase the sensitivity of the detection of bovine milk in ovine and/or caprine cheese by gel isoelectric focusing of γ_2-caseins. Milchwissenschaft 45, 708-711 (1990).

Addeo F., Nicolai M.A., Chianese L., Moio L., Spagna Musso S., Bocca A., Del Giovine L.: A control method to detect bovine milk in ewe and water buffalo cheese using immunoblotting. Milchwissenschaft 50, 83-85 (1995).

Krause I., Berner I, Klostermeyer H.: Sensitive detection of cow milk in ewe and goat milk and cheese by carrier ampholyte — and carrier ampholyte/immobilized pH gradient — isoelectric focusing of γ-caseins using plasmin as signal amplifier. in: Electrophoresis-Forum 89 (B. J. Radola, ed.) pp 389-393, Bode-Verlag, München (1989).

Krause I., Belitz H.-D., Kaiser K.-P.: Nachweis von Kuhmilch in Schaf and Ziegenmilch bzw. -käse durch isoelektrische Fokussierung in harnstoffhaltigen Polyacrylamidgelen. Z. Lebensm. Unters. Forsch. 174, 195-199 (1982).

Radola B.J.: Ultrathin-layer isoelectric focusing in 50-100 µm polyacrylamide gels on silanised glass plates or polyester films. Electrophoresis 1, 43-56 (1980).

Figure 1 **Schematic drawing of the covering sheet**

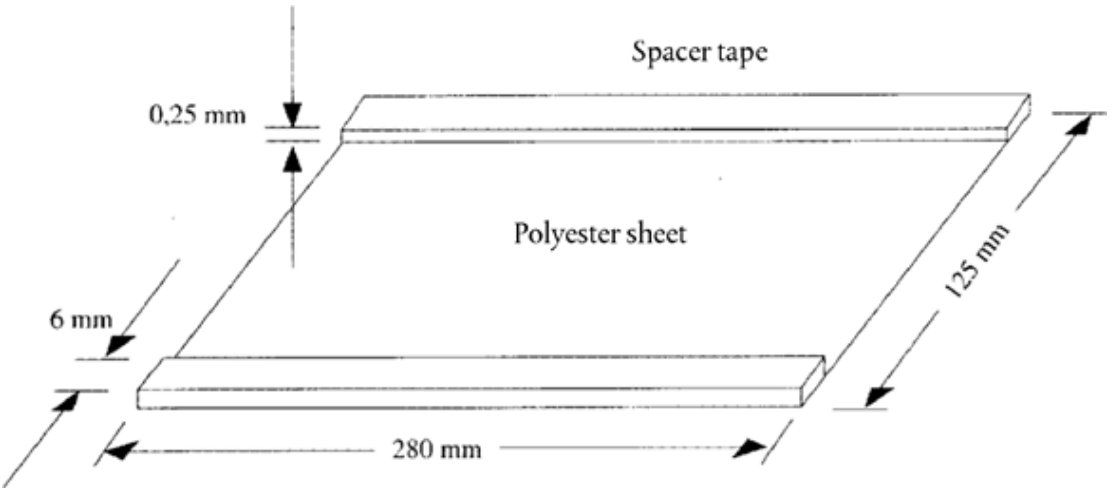


Figure 2 **Casein layer floating between aqueous and organic phases after centrifugation**

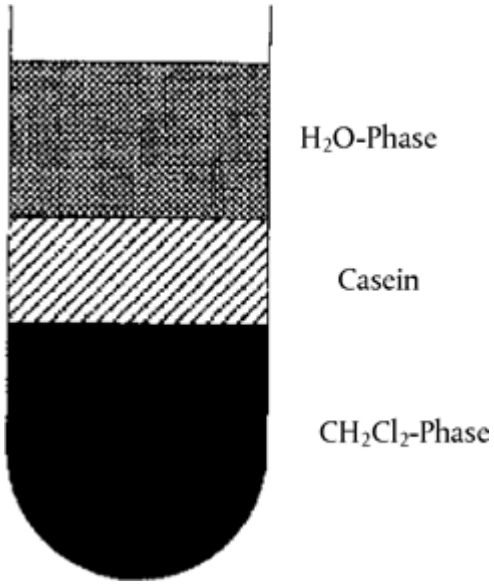
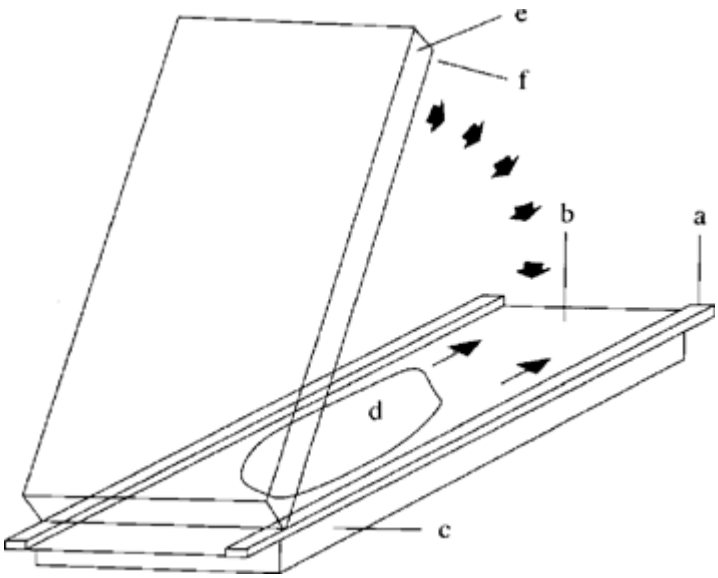
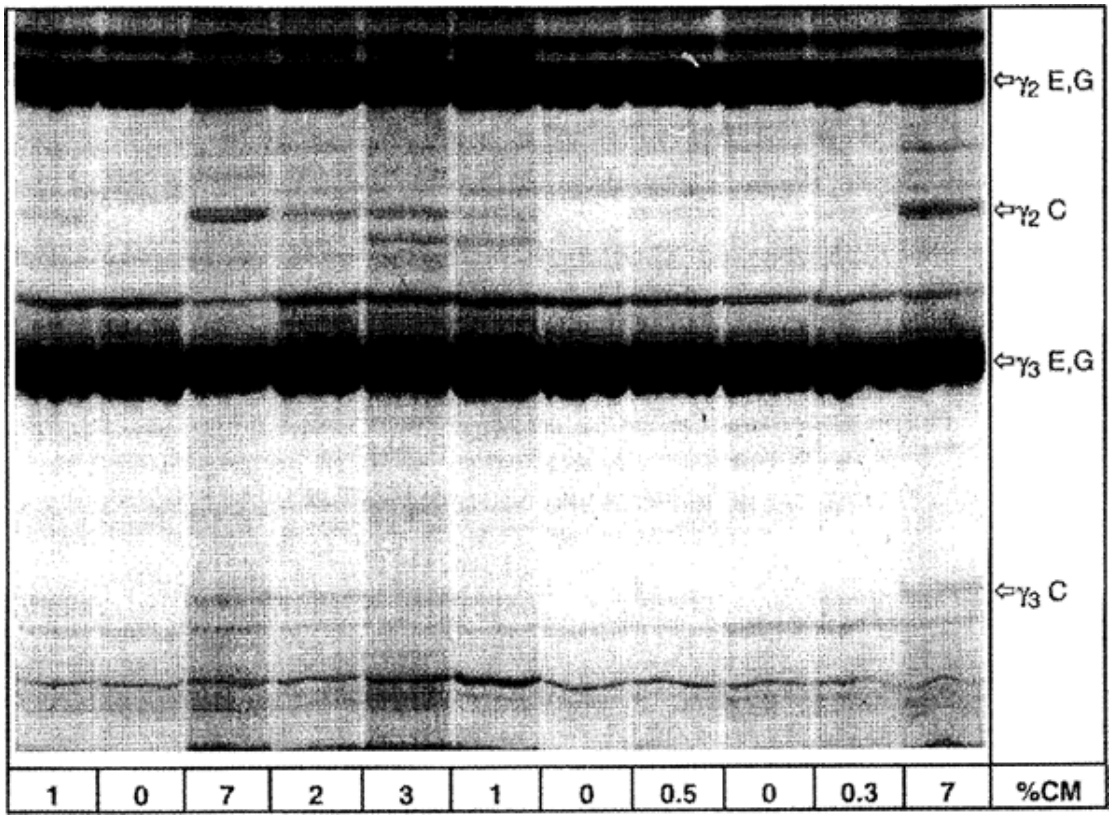


Figure 3 **Flapping technique for casting of ultrathin polyacrylamide gels**



a = spacer tape (0,25 mm); b = covering sheet (5.3); c, e = glass plates (5.1); d = gel solution (4.1.2); f = gel carrier sheet (5.2)

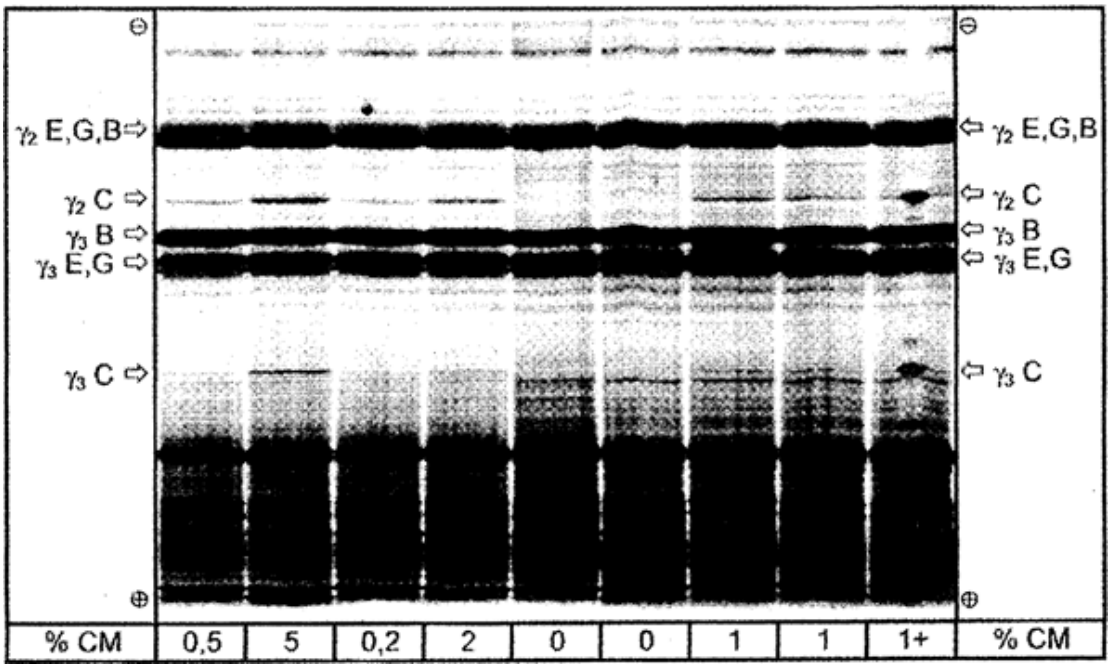
Figure 4a **Isoelectric focusing of plasmin-treated caseins from ewe's and goat's milk cheese containing different amounts of cow's milk.**



% CM = percentage of cow's milk, C = cow, E = ewe, G = goat

Upper half of the IEF gel is shown.

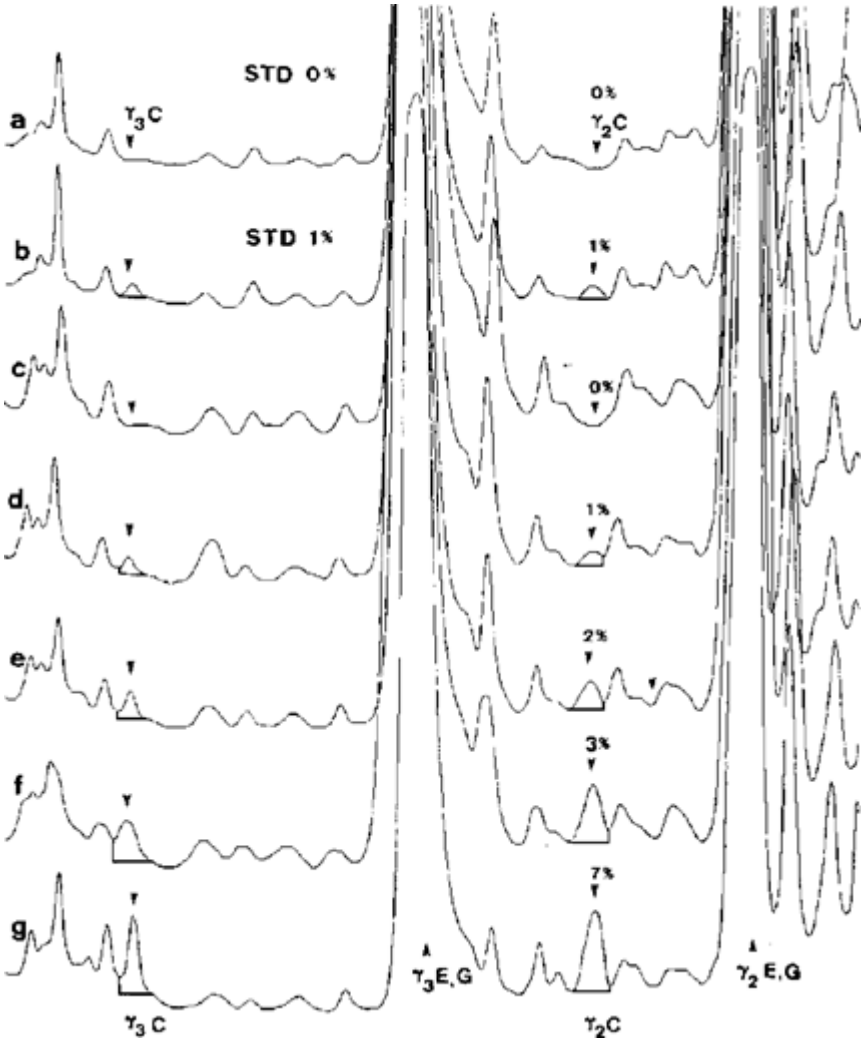
Figure 4b **Isoelectric focusing of plasmin treated caseins from cheese made from mixtures of ewe's, goat's and buffalo milk containing different amounts of cow's milk.**



% CM = percentage of cow's milk; 1 + = sample containing 1 % of cow's milk and spiked with pure bovine casein at the middle of the track. C = cow, E = ewe, G = goat, B = buffalo.

Total separation distance of the IEF gel is shown.

Figure 5 **Superposition of densitograms of standards (STD) and cheese samples made from a mixture of ewe's and goat's milk after isoelectric focusing.**



a,b = standards containing 0 and 1 % of cow's milk; c-g = cheese samples containing 0, 1, 2, 3 and 7 % of cow's milk. C = cow, E = ewe, G = goat.

Upper half of the IEF gel was scanned at λ = 634 nm.**]**

Changes to legislation:
There are currently no known outstanding effects for the Commission Implementing Regulation (EU) 2016/1240.